

# Membrane (and Soluble) Protein Crystallization for Structure-Function Studies Using Lipid Mesophases. *In situ* diffraction data collection

Membrane Structural and Functional  
Biology Group

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*In Situ* Serial Crystallography Workshop

Swiss Light Source, Villigen, Switzerland

*November 17 - 19, 2015*



Nature 477: 549

“This image is a molecular masterpiece ...”

The Royal Swedish Academy  
of Science. October 10, 2012



# Reference Material

- ♣ Slides
- ♣ Acta F71: 3-18. Acta D71:1238-56. Acta D *In press*
- ♣ Nature Protocols 4: 706-31
- ♣ JoVE 45: e1712; 67: e4000; 67: e4001 – Open Access

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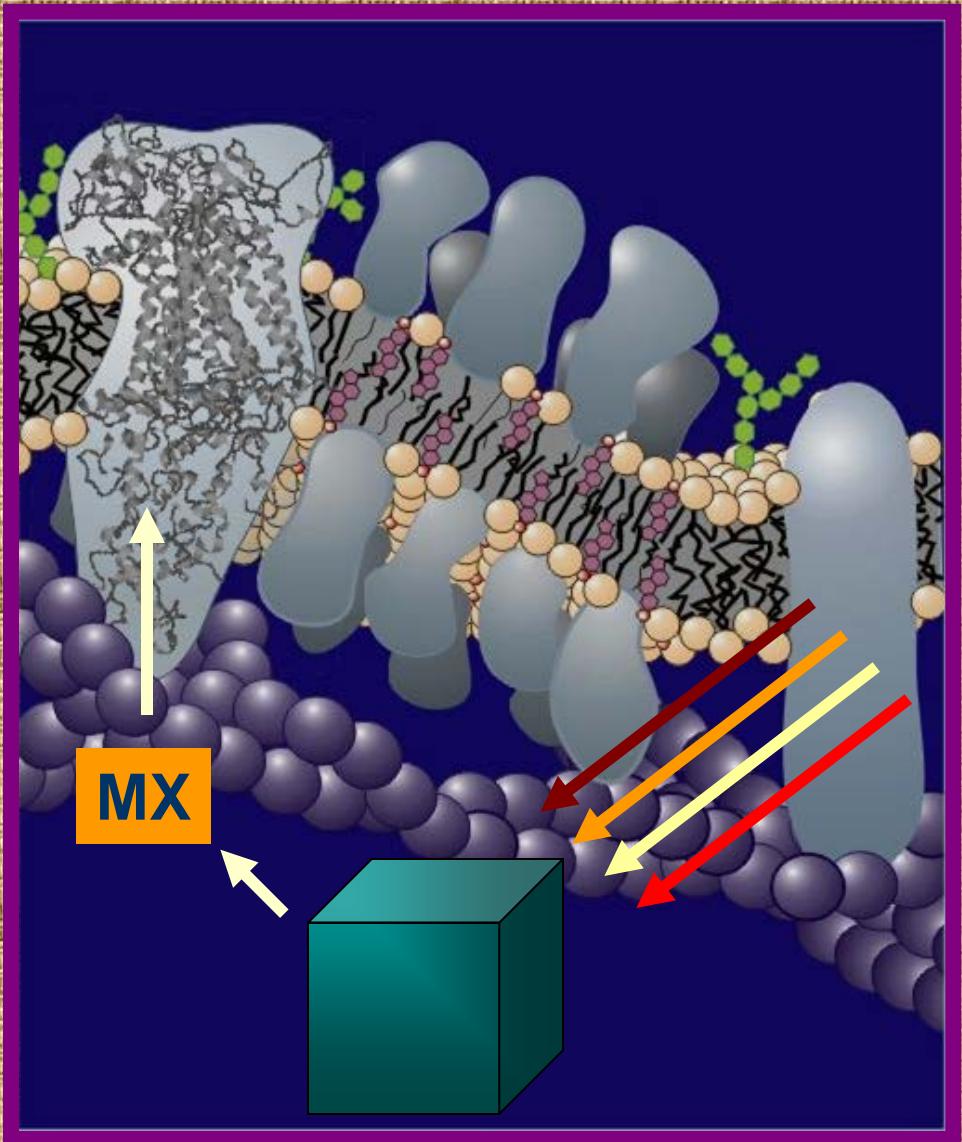
## A fast, simple and robust protocol for growing crystals in the lipidic cubic phase

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A simple and inexpensive protocol for producing crystals in the sticky and viscous mesophase used for membrane protein crystallization by the *in meso* method is described. It provides crystals that appear within 15–30 min of setup at 293 K. The protocol gives the experimenter a convenient way of gaining familiarity and a level of comfort with the lipidic cubic mesophase, which can be daunting as a material when first encountered. Having used the protocol to produce crystals of the test protein, lysozyme, the experimenter can proceed with confidence to apply the method to more valuable membrane (and soluble) protein targets. The glass sandwich plates prepared using this robust protocol can further be used to practice harvesting and snap-cooling of *in meso*-grown crystals, to explore diffraction data collection with mesophase embedded crystals, and for an assortment of quality control and calibration applications when used in combination with a crystallization robot.

# Structural and Functional Biology of Membranes



## Crystallization Methods

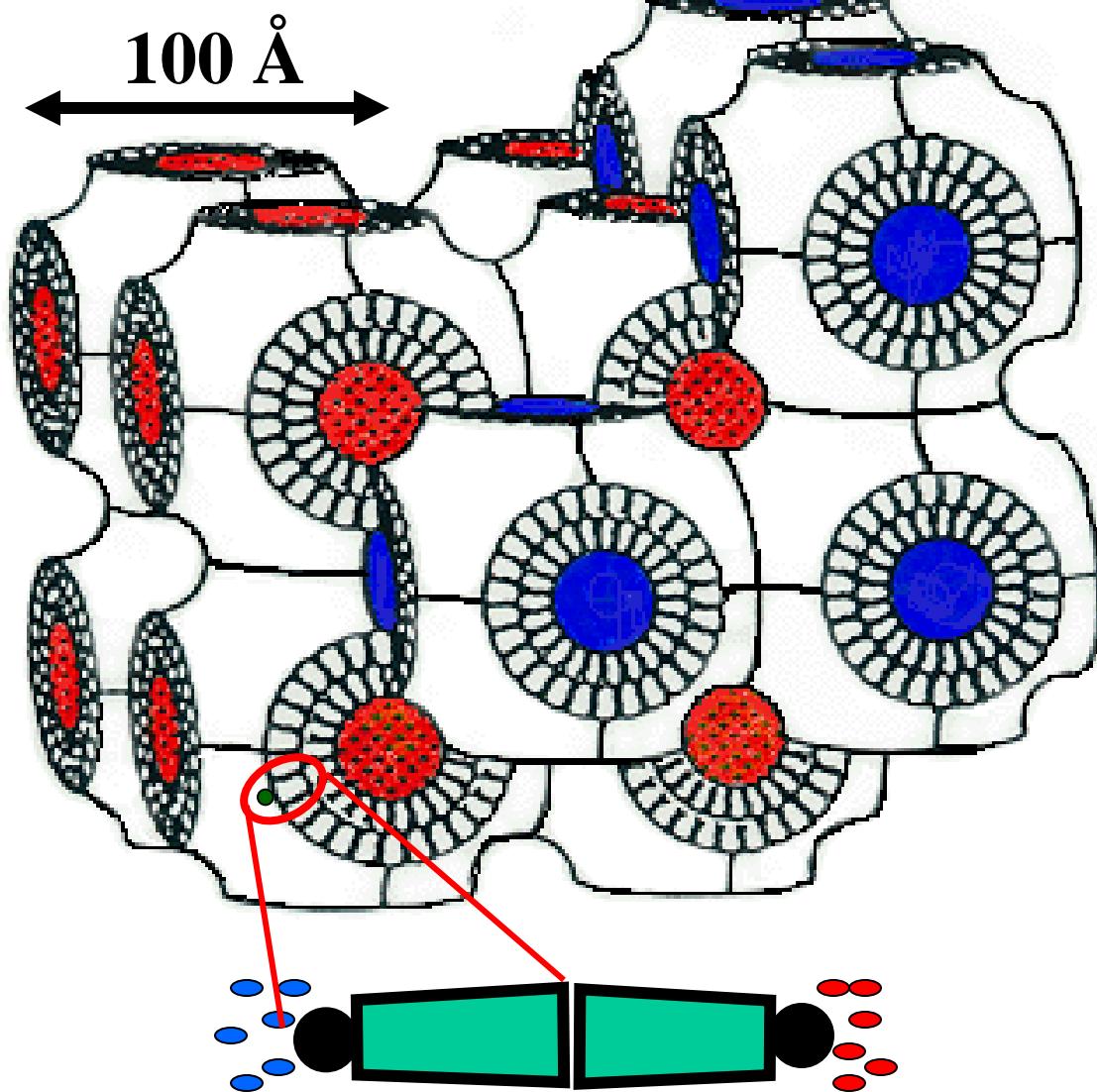
Vapor Diffusion  
hanging drop  
sitting drop

Batch  
Dialysis  
Bilayer Methods

bicelle  
vesicle fusion  
cubic phase

Evaporation  
Interfacial Diffusion

# Cubic Phase



Bacteriorhodopsin crystallized  
in lipidic cubic phase



20  $\mu\text{m}$

Lamellar  
portal

PROTEIN  
CO-CRYSTAL

CHARGE  
SCREENING

Cubic  
phase

Lipids

Crystalline  
array

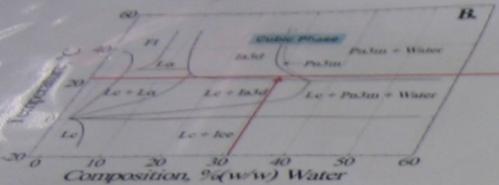
Reconstituted  
protein

J. Struct. Biol.  
142:108-132

Cryst. Growth Des.  
8:4244-4254

♣ Nature Protocols 4:706  
♣ JoVE 45: id 1712

Water MeOH



Chemistry and Physics of Lipids 107 (2000) 191–220

In Miami Decem. 11 1948

Wing 217

Wings posterior margin - top 1/3 moved into mouth & replaced by  
from which finger bracket removed.  $\frac{1}{2}$  front fingered &  
middle & top 2/3 of middle finger bracket joint.  
rest 1/3 of middle finger bracket joint.

Chitons - 1948 - 1950 - Good - and 33% of them  
had to be cut open to see. Sporangia in same way as  
the tip of the alveolar. Other way is to add a vial methylene  
blue to the tip of the alveolar & wait until the spores turn  
blue & then open them - see Cole in 1950 paper.

Spores with 2 furrows on back. Length 25 microns. No finger  
at one & two more added to first three. I just draw  
the tip of the long needle - break off a piece  
so we can hold it up the end & inserted - looks in  
like a willow at first & appears broken off a little.  
- finger height. Use water into liquid



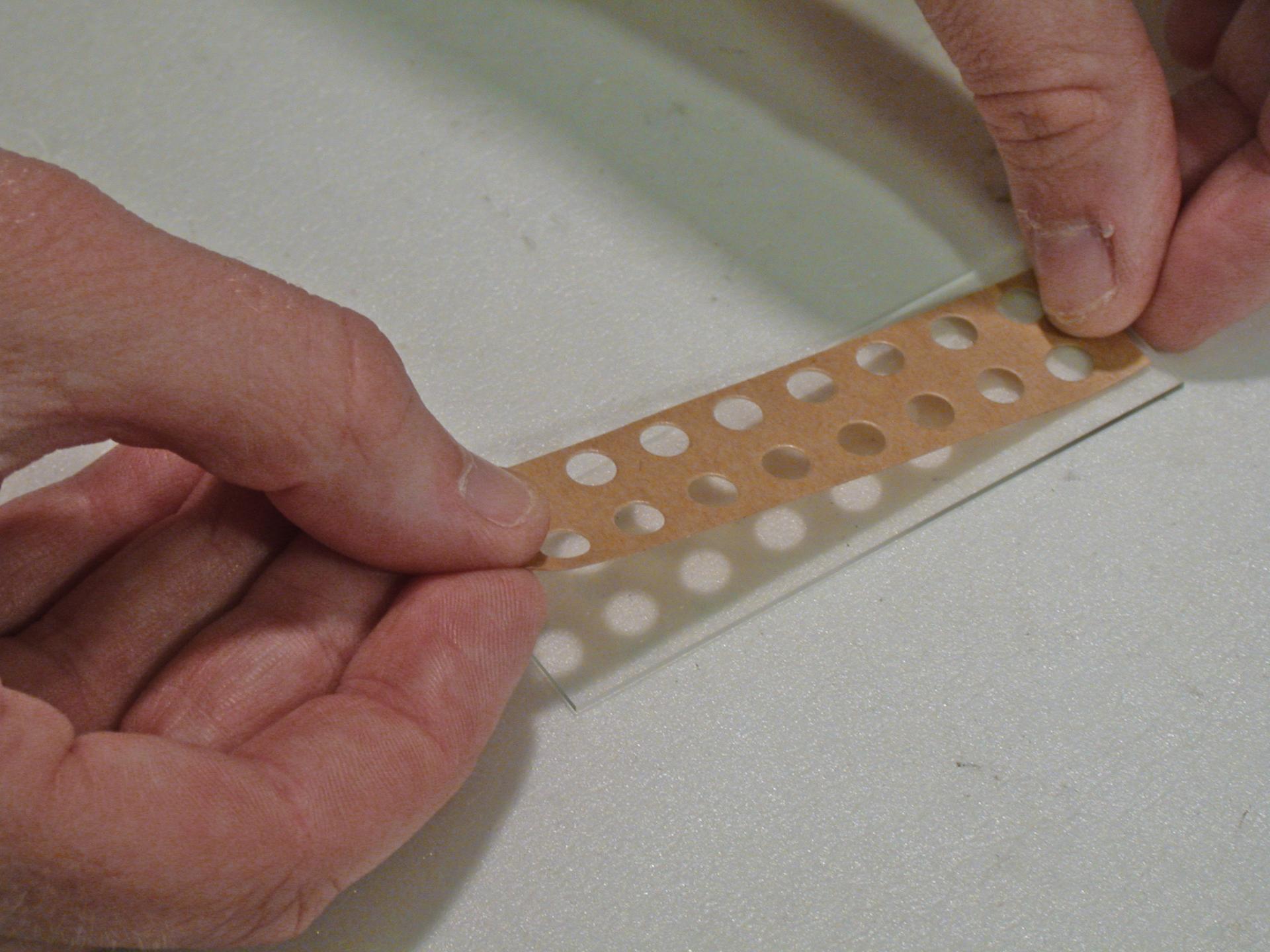
# Calculator

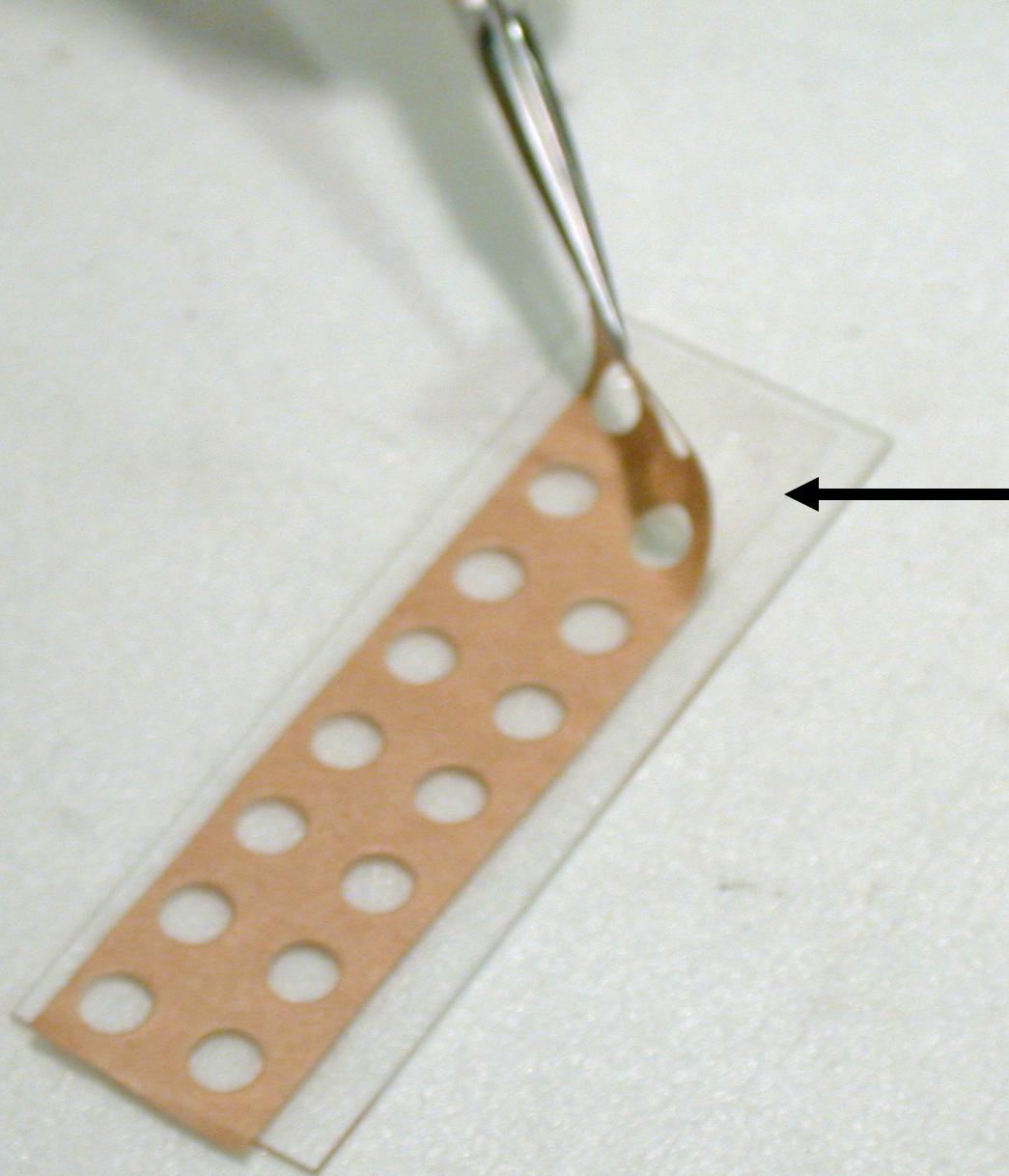
## Ice

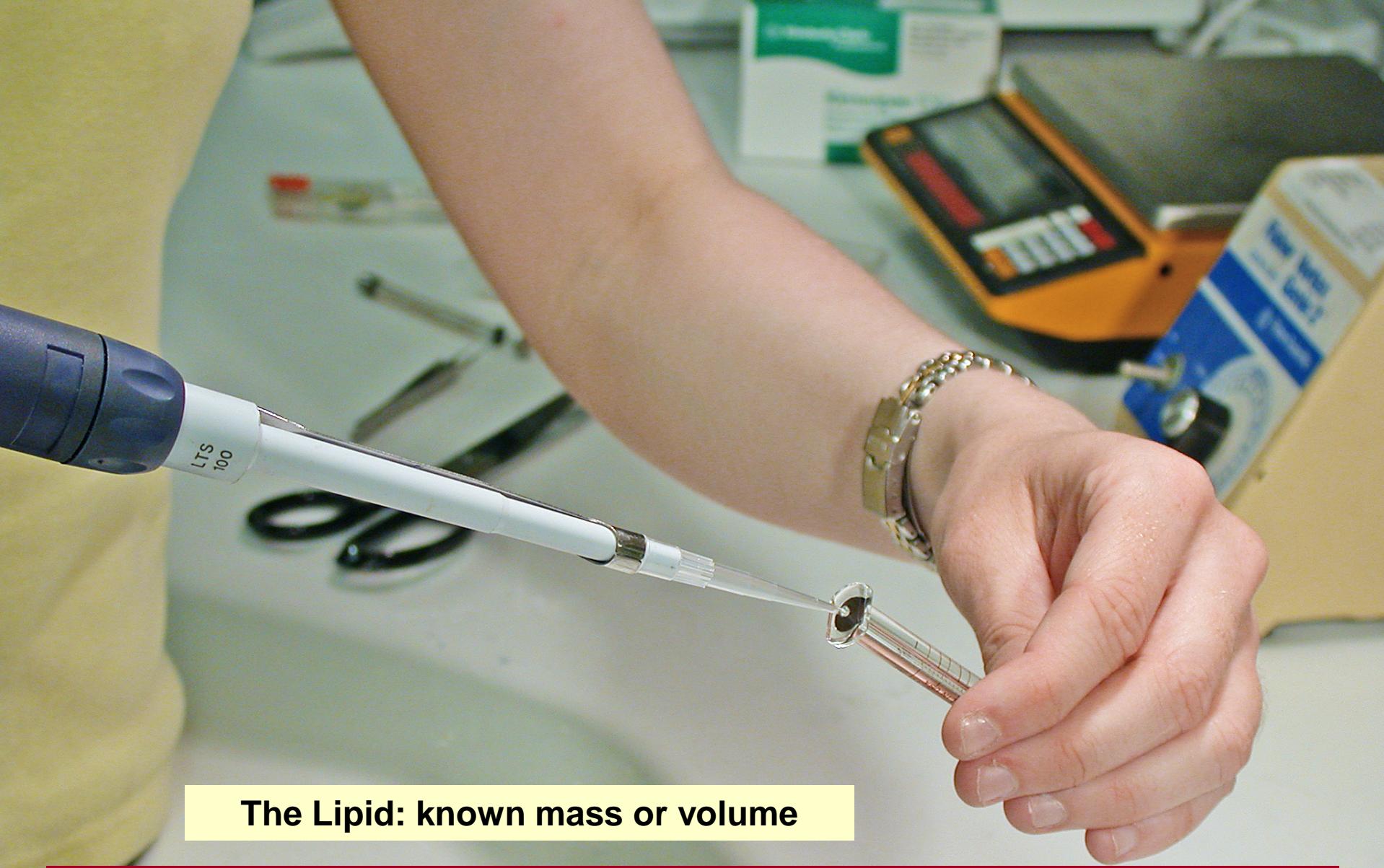
1

30  $\mu$ L







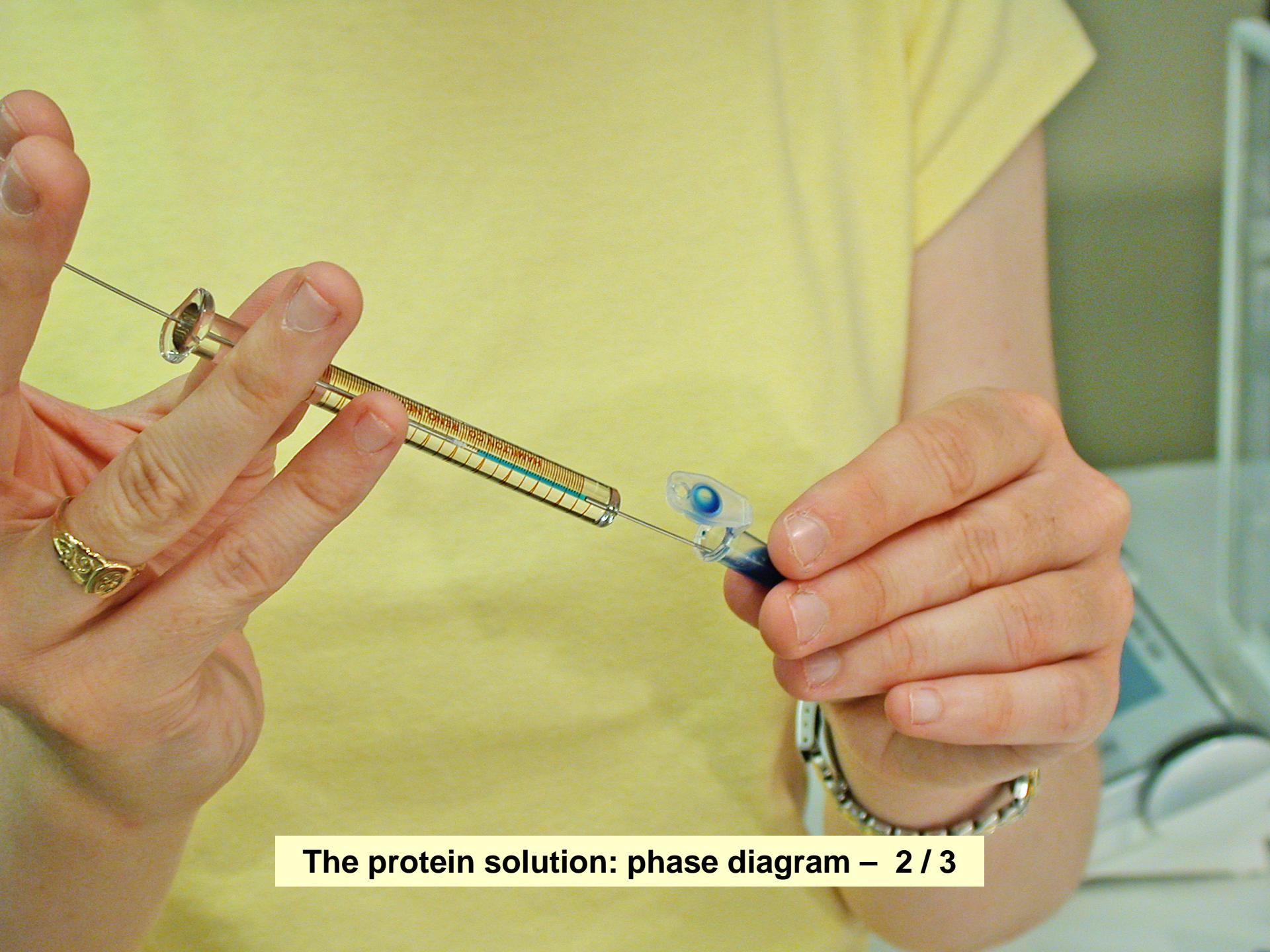


The Lipid: known mass or volume

Lipid

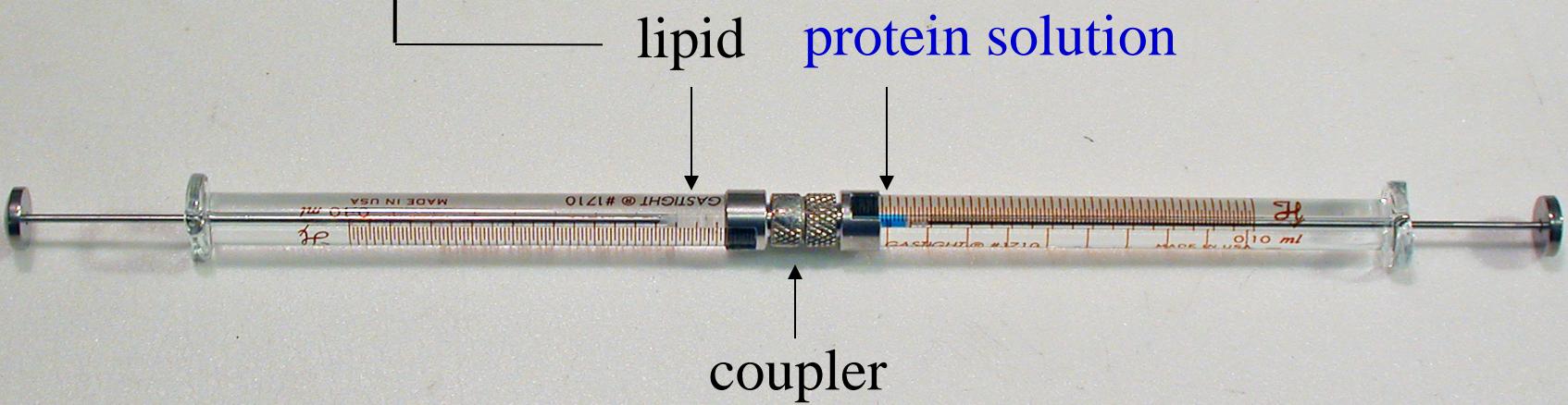
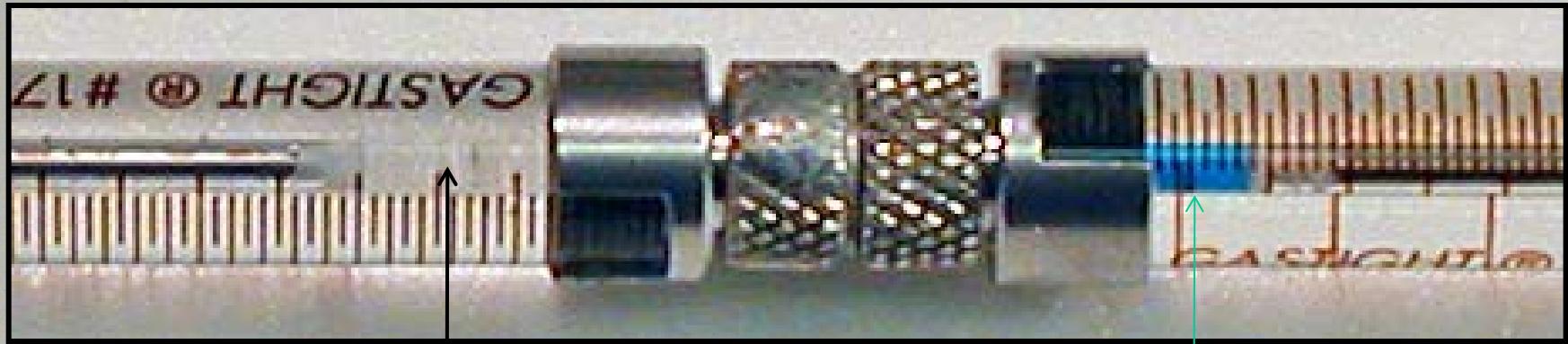
Protein





**The protein solution: phase diagram – 2 / 3**



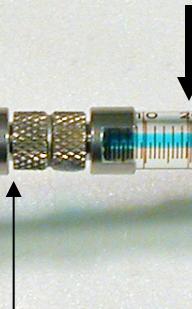


# Coupled Syringe Mixer



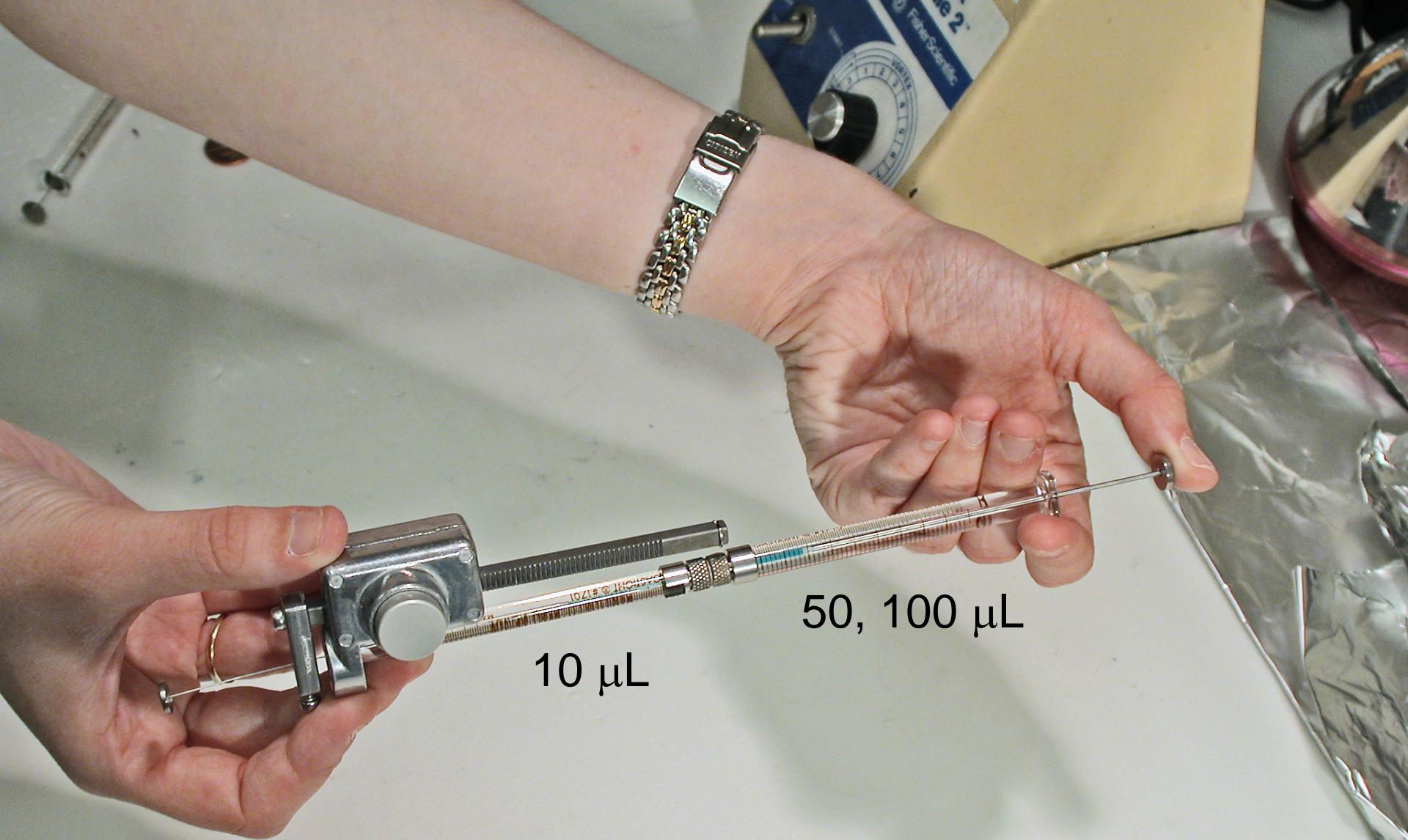
**Mechanically mixing the protein solution and lipid together to form the cubic mesophase**

optically clear  
protein/lipid mesophase



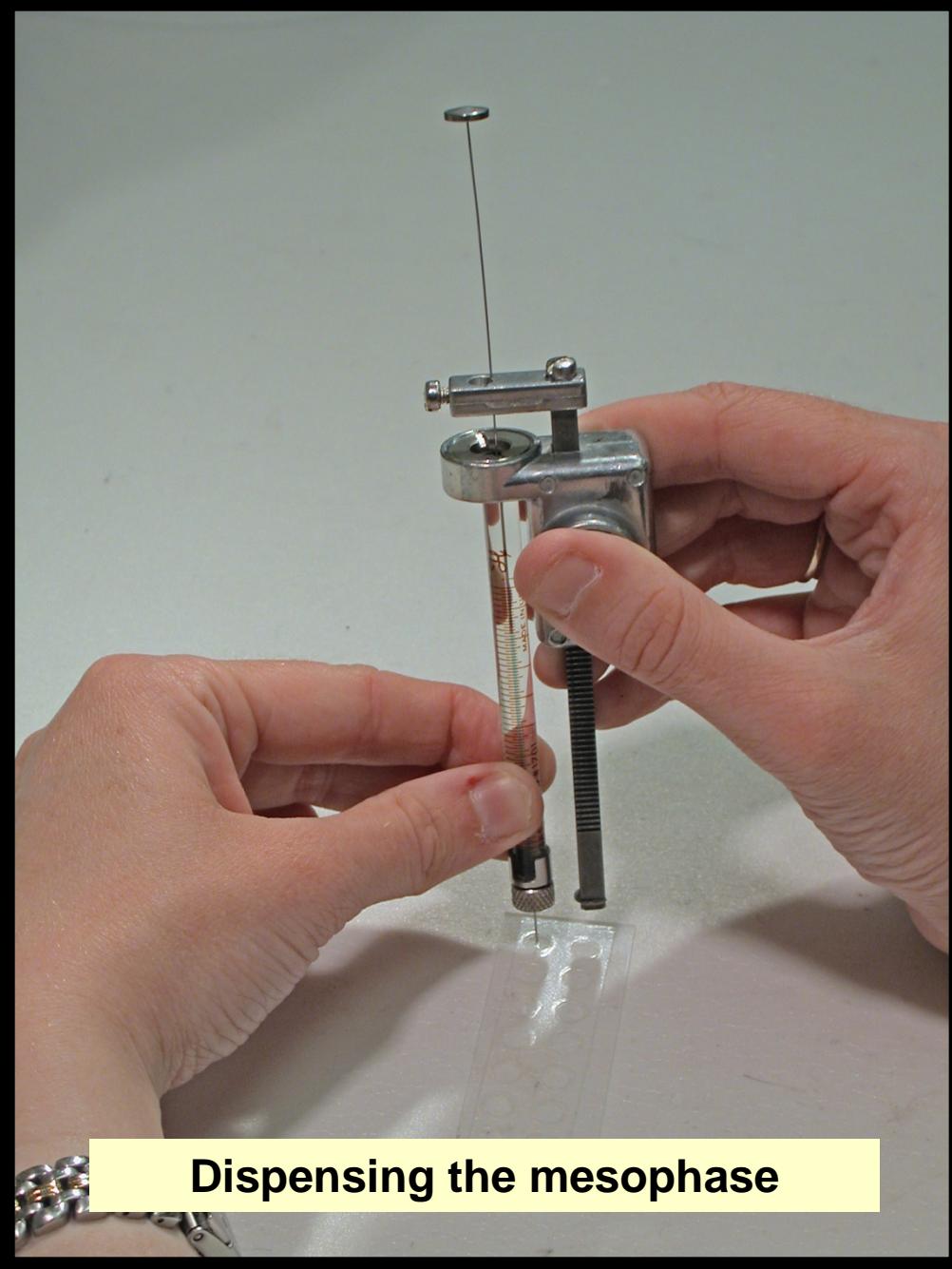
disconnect

The homogenous protein / lipid cubic phase

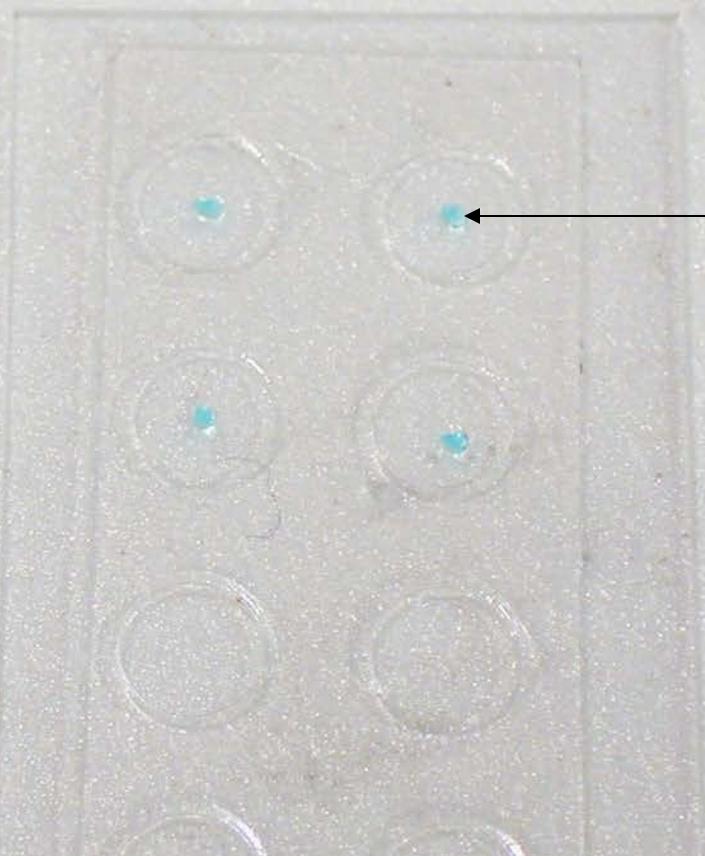


**Loading the dispenser**

10  $\mu\text{L}$   
50, 100  $\mu\text{L}$



**Dispensing the mesophase**



200 nL  
protein/lipid  
mesophase

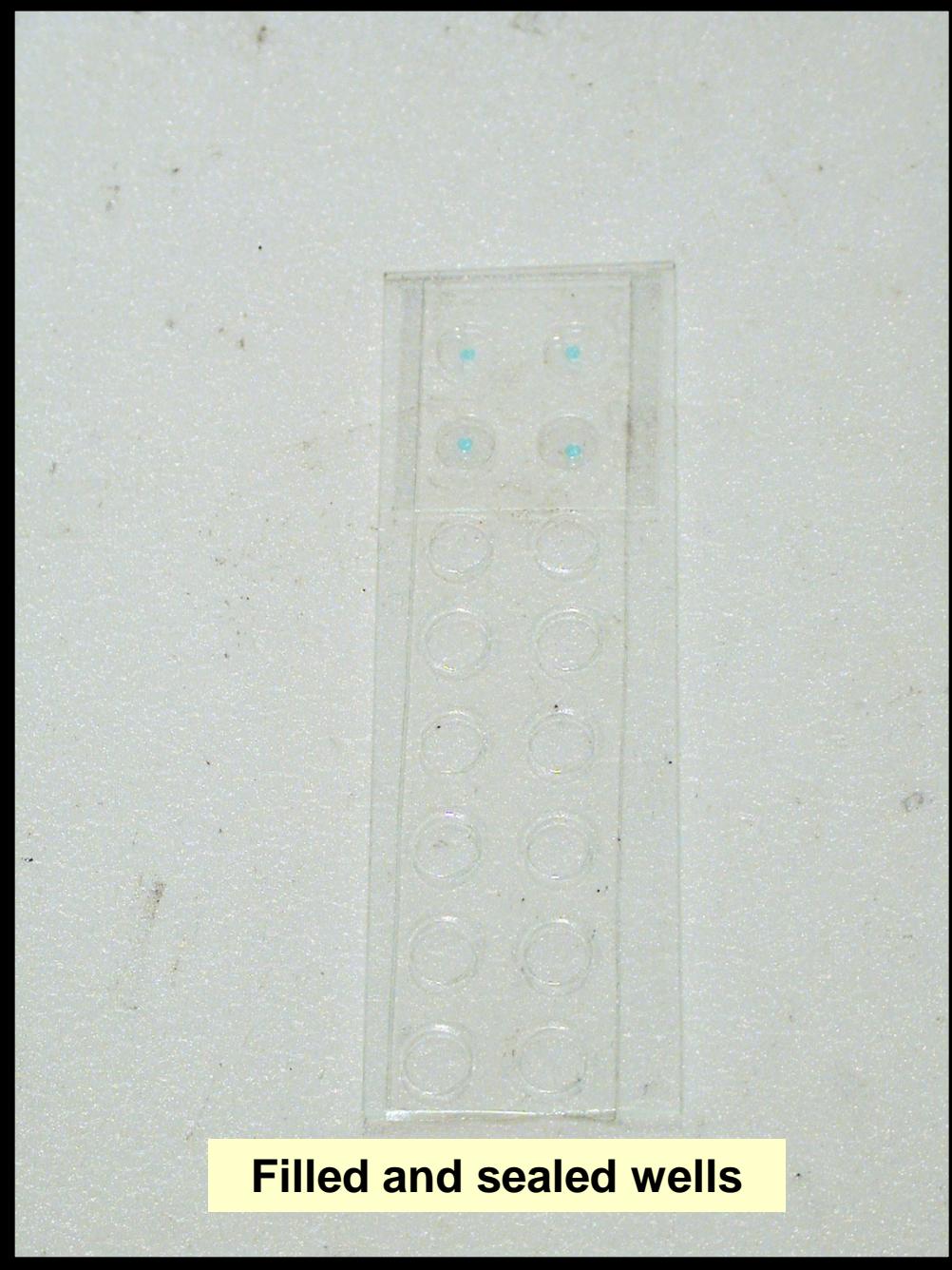
**4 Wells with mesophase**

Dispensing 1  $\mu$ L  
precipitant solution



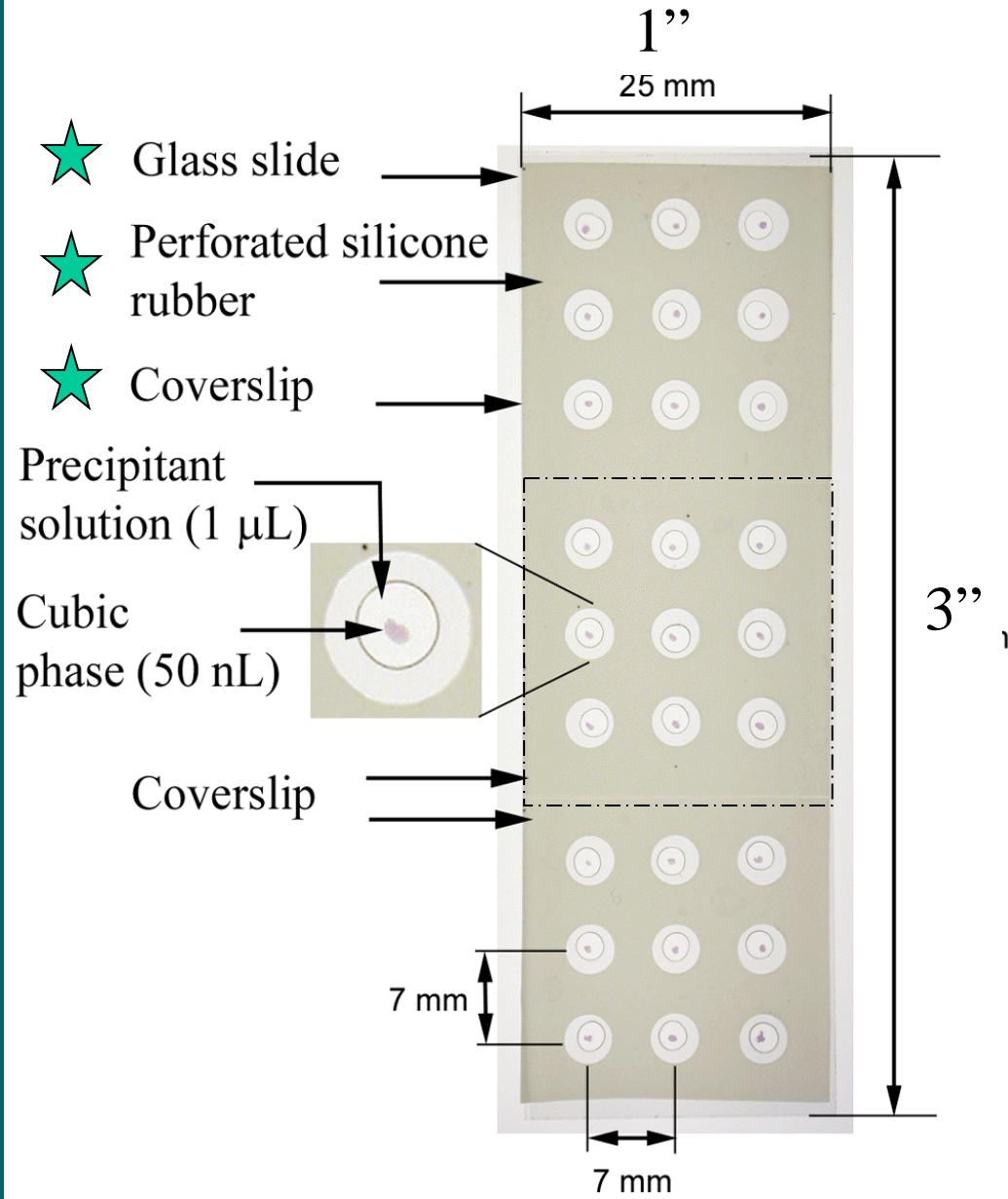


**Sealing wells with glass coverslip**



**Filled and sealed wells**

## Loaded/sealed 27-well crystallization plate



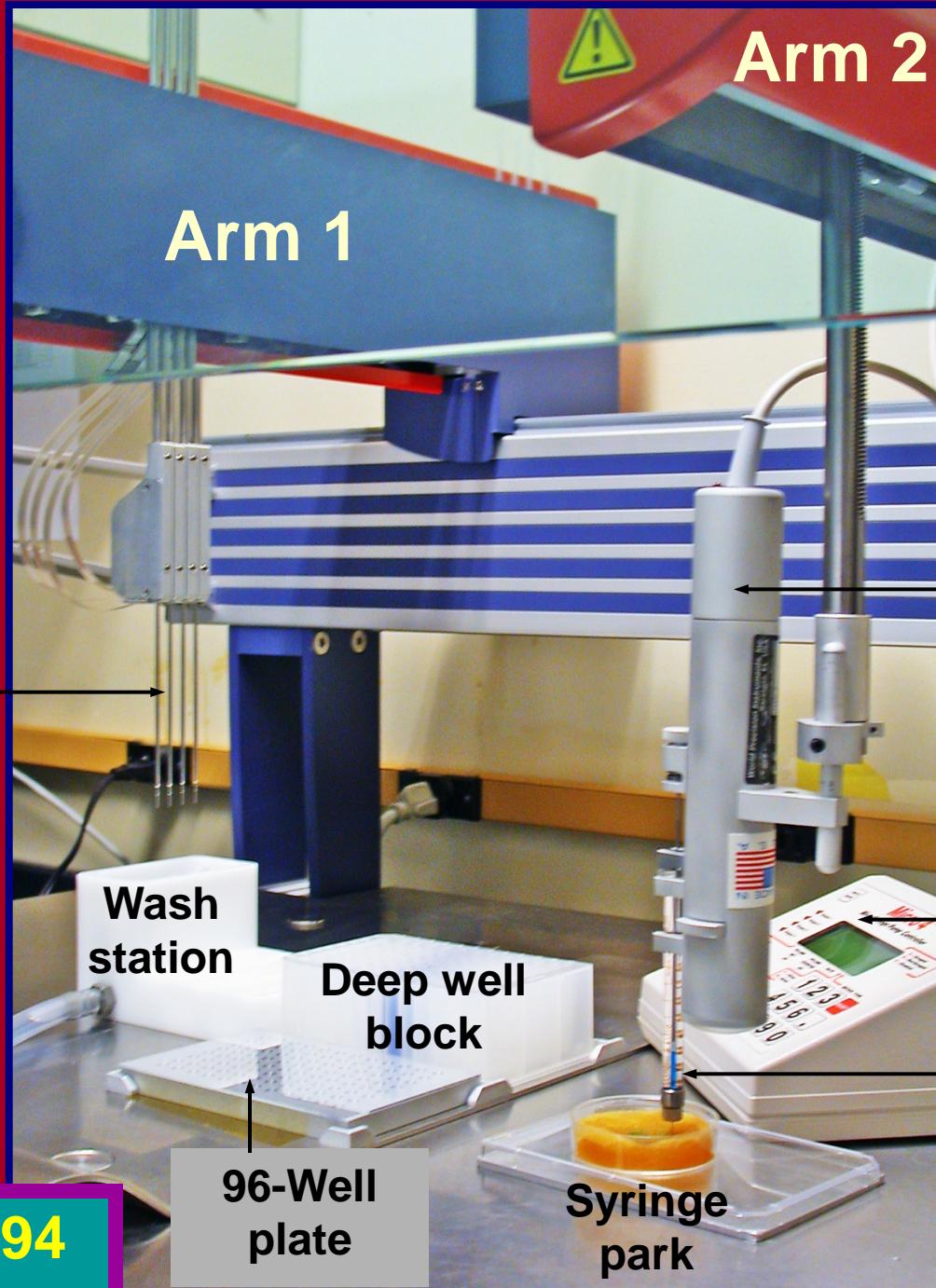
# In Meso Robot

Liquid  
Handling  
Robot

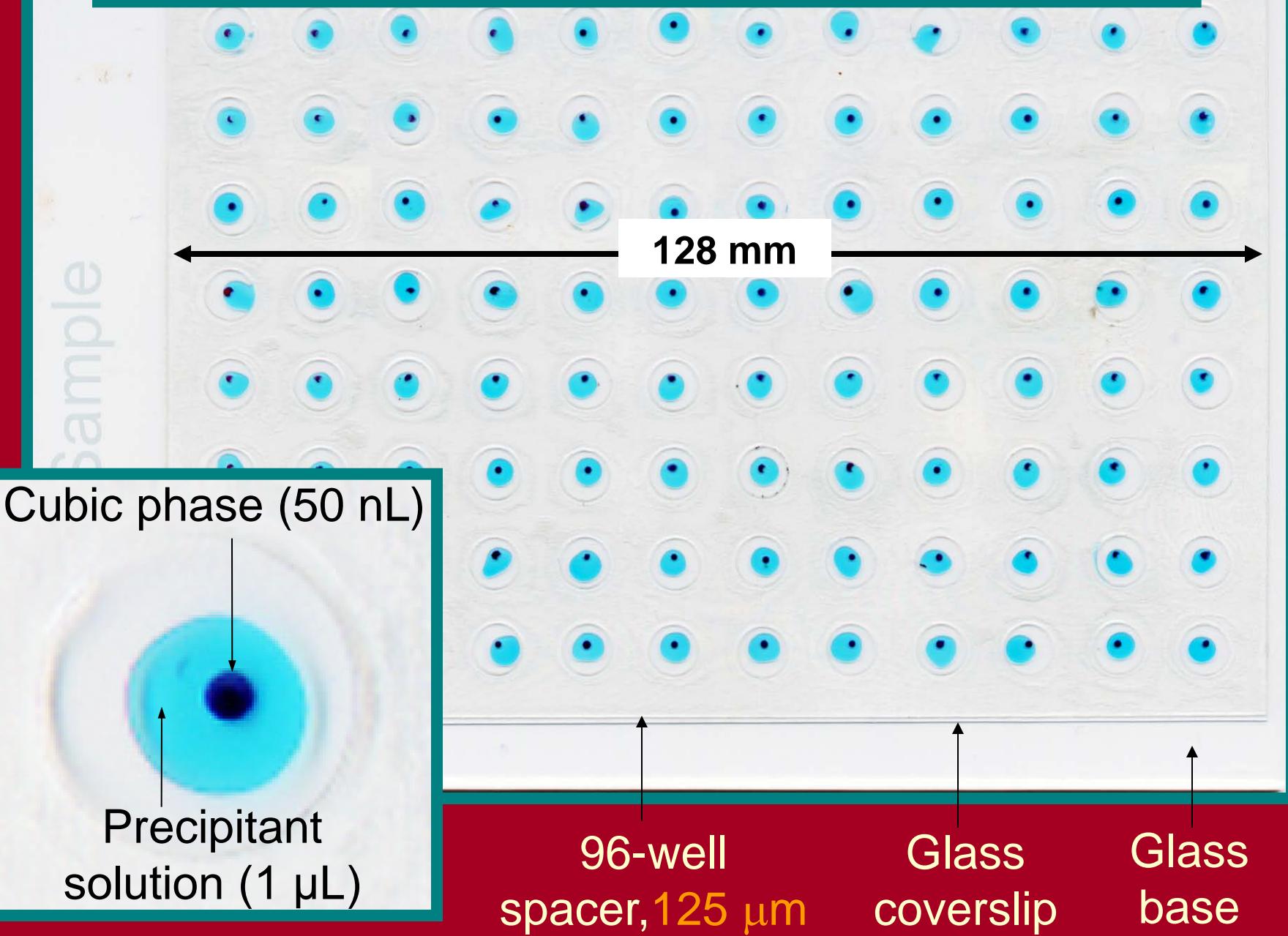
4-Tip  
precipitant  
solution  
dispenser

Motorized  
Micro-pump

Acta D60: 1794

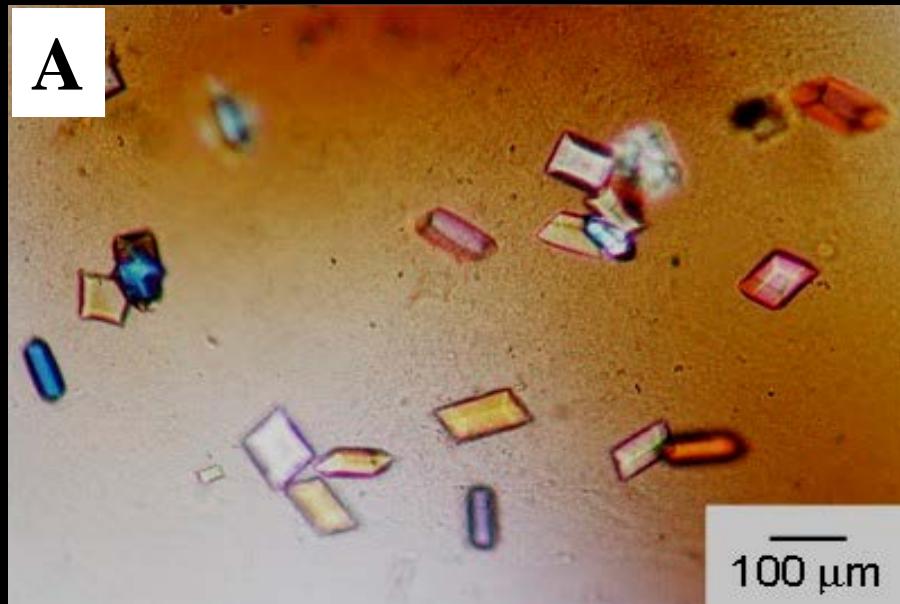


# A Fully Loaded and Sealed 96-well Plate – 5 min

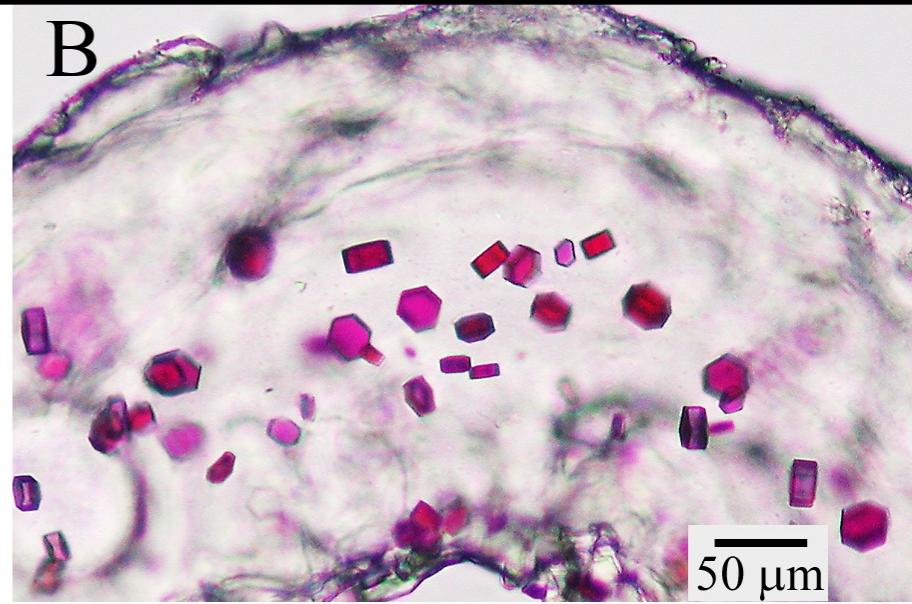


# Crystals $\Rightarrow$ Harvest, Snap Cool, MX

A



B



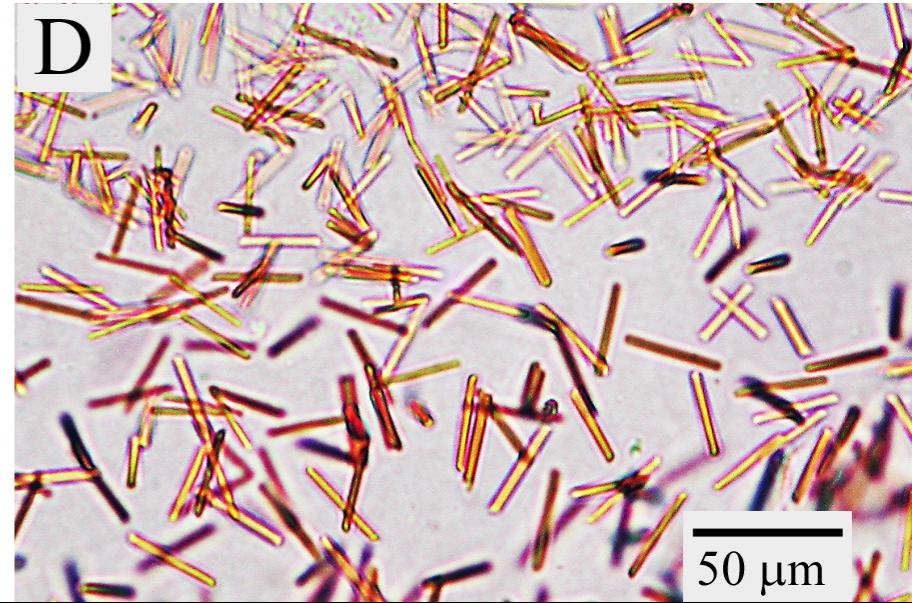
C



♣ JoVE 67: e4001  
(harvest)

50 μm

D

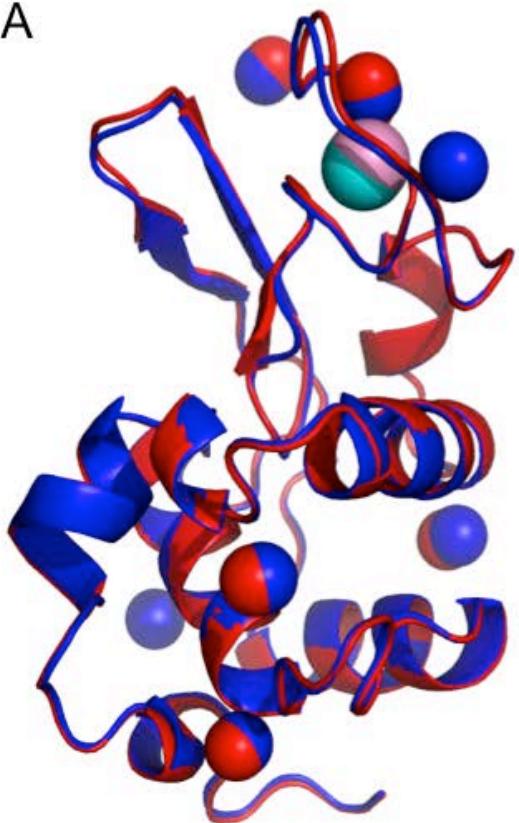


50 μm

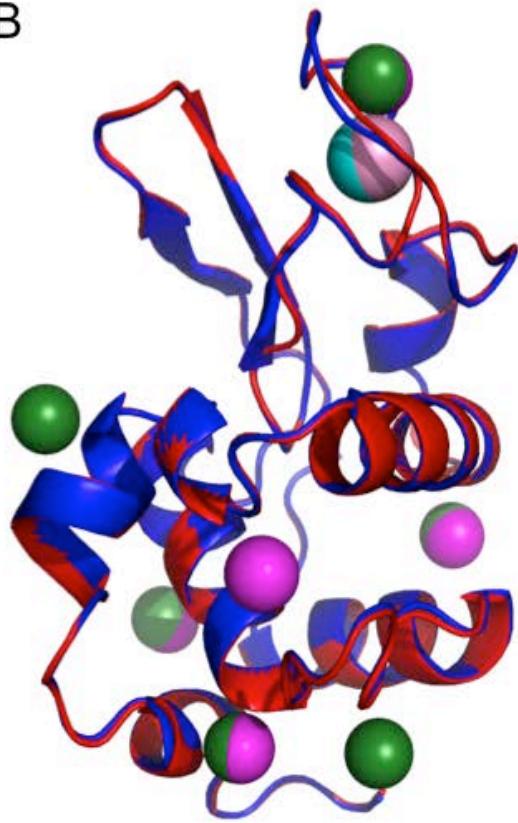
# Water-soluble proteins too !

## Lysozyme

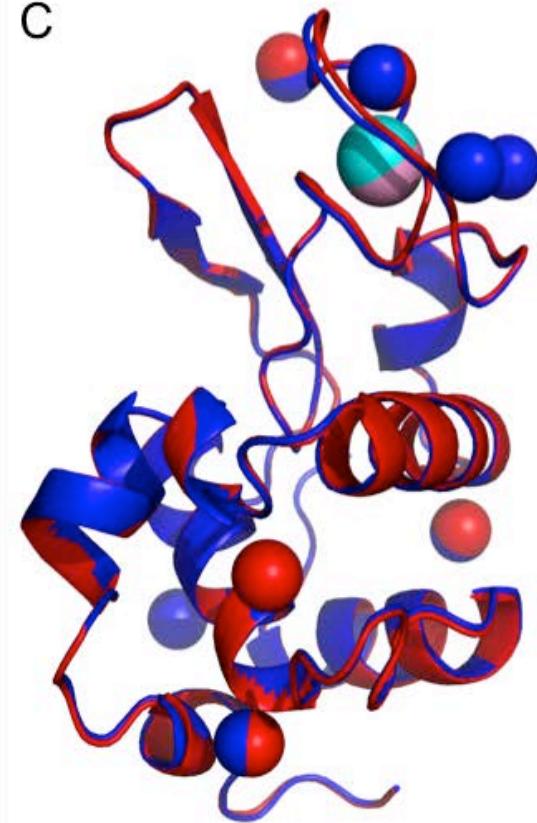
A



B

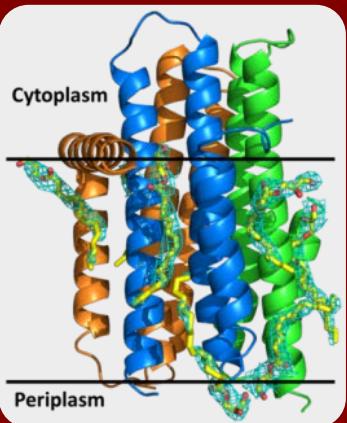


C



+ insulin, thaumatin,  $\alpha$ -lactalbumin, .....

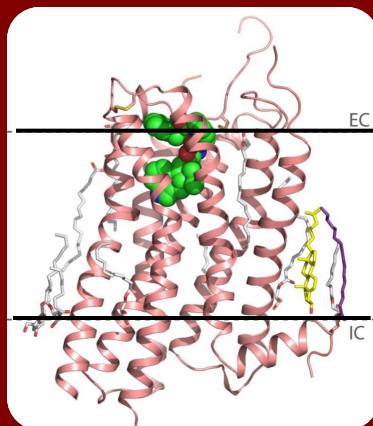
DgkA 2.2 Å



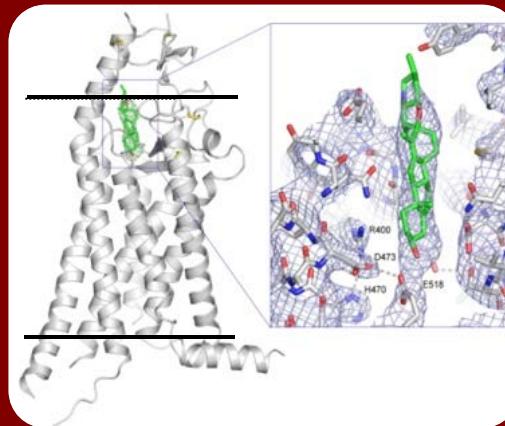
# Jet LCP - SFX Works !

- ♣ Membrane protein structures solved

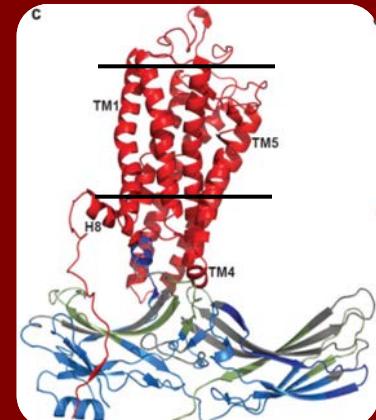
GPCR1 2.8 Å



GPCR2 ~3.2 Å



Rh-Arr ~3.5 Å



♣ Science 342:1521

♣ Phil. Trans. R. Soc. B 369

♣ Nature Commun. 5:3309

♣ Nature 523:561

# *In Meso In Situ SX*

iMiSX

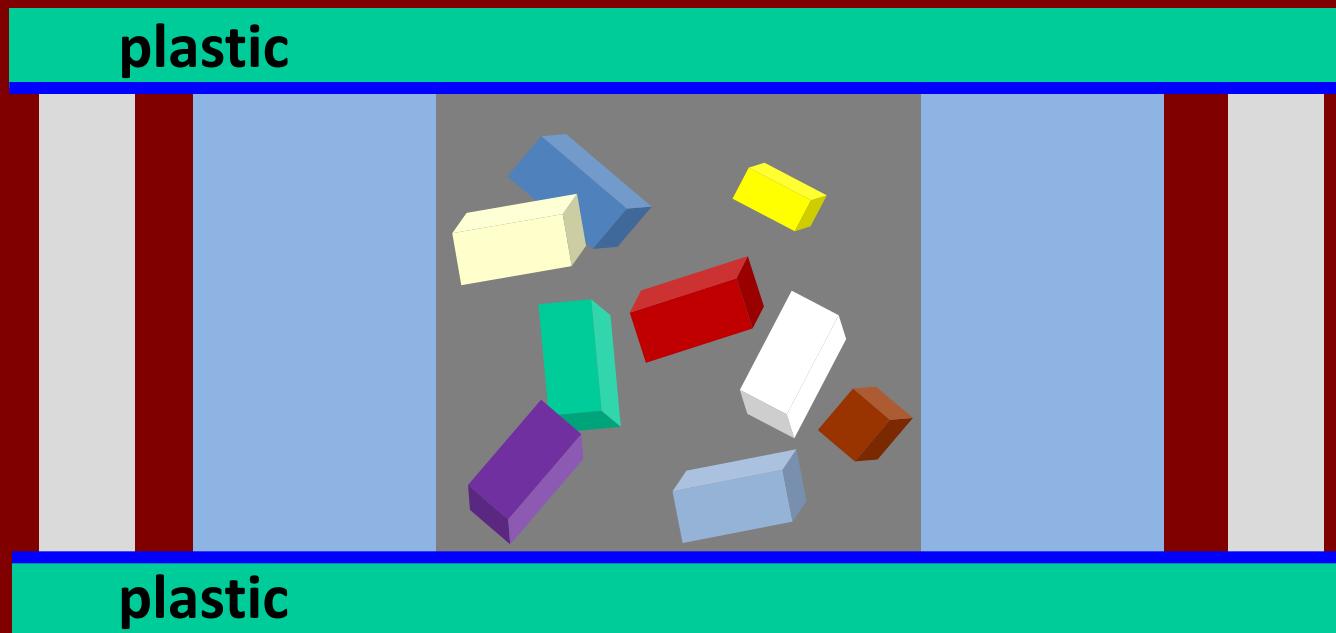
glass

plastic

plastic

glass

Double Sandwich Plate



**Inner Sandwich Plate - RT or Cryo**